



Research Article



The Oxidative Stress Influence in Epigenetic Mechanisms and its Related with Diseases

¹Shaimaa A. Al-Oubaidy, ²Methaq J. Al-Jboori and ³Mona N. Al-Terehi

¹Department of human anatomy and histology/ Medical college/ University of Babylon/Iraq

²Department of biology/College of Science/ Mustansiriyah University/Iraq

³Department of Biology/College of science/ University of Babylon/Iraq

KEY WORDS:

Influence
Oxidative stress
Epigenetic mechanisms

Abstract:

The present review was suggested to explain the effect of oxidative stress (OS) which promote by excessive generation of free radicals like reactive oxygen (ROS) or nitrogen species (RNS) from exo and endogenous factors in the cells in the epigenetic processing's included DNA methylation (DNAM), non-coding RNA and histone modification, these mechanisms regulate gene expression that might be affected in wide range of diseases, oxidative stress impact in epigenetic in different levels included promote nucleotide modification, inhibition or eliminated specific enzymes activity, structural changes of proteins in these pathways, nevertheless, the essential free radicals function in cell processing, it implicated in up and/or down regulation of some gene expression via epigenetic alterations. significant contribution of OS in epigenetic changes via different mechanism, particularly in absence of antioxidant defense, that effect in cell components, changes in some physiological markers like hormones, enzymes, regulatory factors lead to tissues changes and organ dysfunction.

Corresponding Author:

Shaimaa A. Al-Oubaidy,
Department of human anatomy and histology/ Medical college/ University of Babylon/Iraq

INTRODUCTION

Changes of histones, DNA sequences and non-coding RNAs, in addition to DNA sequence changes, Which result to the chromatin remodeling to enhance or obstruct level of gene expression. At the chromatin level the gene expression silencing is essential to the natural life of eukaryotic organisms and it is essential in the vital

mechanisms controlling like genomic imprinting, embryonic development and differentiation^[1], three main mechanisms of epigenetic regulation are demonstrated below:

DNA Sequences Methylation: One of the well-studied mechanisms in epigenetic is DNA methylation (DNAM).

Generally, it is genic silencing synonym, because it shapes of chromatin as an inaccessible state for the transcription process. In DNAM methyl group (CH₃) was addition to carbon 5 of a cytosine, in general, regions rich with 5'-Cytosine-phosphate-Guanine-3' (CpG) dinucleotides found in asymmetric pattern in the genome and this motifs in the promoter regions of gene is known as CpG Island (CGI)^[2,3]. About 30000 unmethylated CGIs in the human genome, that warrants the potentially active constitutive genes configuration. The motif of methylation is involved in cell differentiation., therefore the methylation dysregulation lead to several diseases^[4]. DNA-Methyltransferases (DNMTs) catalyzed DNAM process, There are four kinds of DNMTs, DNMT3A and 3B contributed in de-novo DNAM through development, DNMT1 contributed in methylation patterns maintaining after replication of DNA and DNMTs represented in accompanied with de novo to induce chromatin remodeling complexes without catalytic site which is important in embryonic development^[3,5]. This process is associated with the dosage compensation in mammals and genomic imprinting^[6-9]. In spite of this silencing is made by DNAM, it is an important to use whole epigenetic machinery to correct process development.

Histone Modifications: The second epigenetic mechanism is Histone Modifications based on the chemical modification in chromatin. As well as of other proteins, posttranslational alterations involvement in proteins processing like acetylation, methylation, ribosylation, phosphorylation, sumoylation and ubiquitination^[10,11]. Whose major function is DNA accessibility regulation, In general ribosylation and phosphorylation stimulate transcription of gene and euchromatin, while sumoylation promotes gene silencing and ubiquitination can has a dual role^[12-21].

Non-Coding RNAs: A new demonstrates groups of epigenetic are regulators Non-coding RNAs (ncRNAs) that can control on gene expression without DNA sequence changes. Before ncRNAs discover, it considers as non-benefit sequences accumulated during evolution^[22]. Nevertheless, the DNA Elements project Encyclopedia involved in the non-coding RNAs classification with known functions also detection of novel ncRNAs^[23-25]. furthermore, ncRNAs are categorized into long ncRNAs (lncRNAs) and small ncRNAs which categorized to microRNAs (miRNAs), P-element Induced Wimpy-interfering RNAs (piRNAs) and small interfering RNAs (siRNAs).

The first epigenetics concept was suggested by

Waddington in 1939, who reported "the actual interplaying between genes and their products to phenotype into being"^[26], Later further explanation was updating, Holliday^[27] and Deans and Maggert^[28] proved that epigenetic is gene expression regulation mechanisms without alteration in DNA sequences. These regulations are implemented via different pathways like DNAM, histone modifications and noncoding RNAs^[29]. Reports have been proved that these processes are essential in the regulation of gene expression in the tissue, genomic imprinting and inactivation of X-chromatin. All these mechanisms work together to constructed "the epigenetic landscape" to demonstrate unique features and patterns division in cell types^[30].

The first epigenetic mechanism that well understood is DNAM, although of DNAM is a stable epigenetic symbol, which may be token off resulted from passive or active demethylation activities. Passive demethylation can be implemented via multiple DNA replication cycles in the lack of methylation functional enzymes^[31], DNMT enzymes downregulation^[32]. DNMT cytologic localization^[33] and DNMT recruitment impairment on DNA^[34]. The 5mC elimination has been implemented via of 5-hydroxymethylcytosine formation, oxidized via the ten-eleven translocation (TET) enzymes. further processed were happened by thymine DNA glycosides forming a site would be repaired by the BER system or deaminated by AID deaminated produce 5-hydroxymethyl uracil (5hmU), that removed by the TDG^[35,36].

Oxidative Stress (OS): the redox state disturbance that proved implicated in different disease Oxidative stress is known as OS, the disequilibrium between free radicals included ROS or RNS and the antioxidants defenses mechanisms in the biological system^[37]. ROS are higher activity molecules including different chemical species, like hydrogen peroxide (H₂O₂), superoxide anion (O₂⁻) and hydroxyl radical (OH). These active molecules can be interaction with cell components as well as proteins, lipids and DNA, RNA cause metabolic pathway alteration, molecular damage which implicated in different diseases pathogenesis^[38]. The main source of ROS is mitochondria resulted from electron transfer via ATP generation^[39,40]. High level of ROS also generated from exogenous factors like ultraviolet irradiation and radioactivity. Cell used antioxidant defense mechanisms to prevent harmful effects of OS., these mechanisms are including enzymatic and non-enzymatic pathways. As well as glutathione

peroxidase, glutathione-S-transferase P1, superoxide dismutase, sulfiredoxin, peroxiredoxin and catalase^[41]. the non-enzymatic molecules consist of vitamins C, E, A and glutathione which have low molecular weight^[42].

The Correlation Between Oxidative Stress and Epigenetics: Oxidative stress was happened as a results of ROS accumulation., evidences found that its increased with age and it is accompanied with cell repair machinery decline, which will causes high level of DNA lesions lead to mutations like in the epigenetic state disruption. several examples of tight interconnection interplays between the OS effects and the epigenetic landscape are discussed below, as well as ROS can effect on the methylome via the oxidized DNA damage formation by pyrimidine and 5-methyl cytosine (5mC) hydrolyzation, regarding to similar structure with the epigenetic signals associated with 5-hmC that can be interfered^[43]. the DNA demethylation can be affected also via TET-mediated hydroxymethylation and DNA oxidation^[44]. Indirectly, ROS can change the epigenetic system activity because histone-modifying enzymes related to intracellular essential metabolites levels like Fe, Acetyl-CoA, ketoglutarate, NAD+and S-adenosylmethionine, proving that the alteration in epigenetic are strong associated with global energy levels and cellular metabolism the cell^[45], Thus OS can globally effect on the cell more than one levels, like DNA and histone modifiers, that will impact in the epigenetic landscape directly.

Oxidative Stress Effect on DNA Methylation: The DNA methylation alterations are generated from OS mainly according to the function and activity of DNMTs changes. Numerous reports in cancer proposed that the OS induction mediated by H₂O₂ raised DNMT1 activity and linking to the tumor suppressor genes promoters^[46]. Furthermore, OH stimulate a global hypomethylation according to the interference of DNMTs-DNA linking capacity^[47]. it has been found that high OS state stimulate catalytic cycle changes of iron and inhibition TET family DNA demethylases, thus DNAM levels was increasing^[48]. More cited studies indicated that high OS situations stimulated via ROS generation increase levels of 8-hydroxydeoxyguanosine (8-OHdG) which promote a conformational modification which change the state of chromatin from active to repressive state^[49]. furthermore, 8-OHdG prevent DNMTs-DNA binding, results to a global genome hypomethylation^[50,51].

Additionally to the directly DNA modification , ROS also impact DNAM pattern, that associated with

transcription repression, the primary methylation In mammals are targeting cytosines in CpG dinucleotides. In spite of CpGs are methylated genome-wide, regions with high density of CpG dinucleotides are especially hypomethylated^[52]. the category of DNAM enzymes included DNMTs and the demethylases, DNMTs used S-adenosylmethionine (SAM) to methyl group transfer to C, while TETs oxidize 5-methyl cytosine (5-mC) to 5-hydroxymethyl cytosine (5-hmC) in an Fe(II)-, ascorbate, O₂-dependent reaction and a-ketoglutarate^[53,54].

In some disease pathologies global DNA hypomethylation has demonstrated, most islands (CpG) are hypermethylated^[55-58], these methylation is changed included specific hypermethylation and global hypomethylation which have been proposed to be related to oxidative stress.

However, the direct 5-mC oxidation to form 5-hmC cans consequence to DNA demethylation. 5-hmC prevents maintenance of Methyltransferases DNMT1, therefore, suitable inheritance of methylation motif blocking and caused indirect demethylation of CpG region^[59].

An approach of DNA hypermethylation under OS is TET proteins inhibition. As Fe(II)- and a-KG-dependent dioxygenases, it can be inhibition by iron oxidation. Through the catalytic cycle, Fe(III) and Fe(IV) is oxidized from Fe(II)^[54] which regenerated by ascorbate^[53]. In condition of OS, this regeneration is concerned, consequence to enzyme inhibition and global 5-mC levels increasing^[60]. Likewise, 5-hmC is important eliminated in SY5Y neuroblastoma cells exposure to OS^[61]. Moreover, in murine cerebellar granule cells, TET1 knockdown lead to cell death, reporting that OS caused TET1 inhibition could be involved in the pathogenesis of some disease^[62].

Both methylation patterns., global hypomethylation and CpG islands hypermethylation can be implemented by DNMTs relocalization. The repair proteins discriminate 8-oxo-dG and induce DNA Methyltransferases DNMT1 maintenance, that made a structure with the de-novo DNA Methyltransferases DNMT3B, the histone H3K27 Methyltransferases and EZH2 SIRT1 deacetylase^[63,64], which is mainly induce CpG island promoter regions activity, when present high level of 8-oxo-dG and DNAM and H3K27me3 enhancing H3K4me3 and H4K16ac levels were reducing. Take together, DNMT3B, DNMT1 and SIRT1 are missed from other genomic location. thus, low expressed genes CpG islands were hyper methylated in an in vivo under OS exposure, while housekeeping genes were unaffected^[63].

Effect Over Histone Modifications: Histones are extensively changed when exposure to free radicals and

are glutathionylated in a redox-sensitive way, that impact its stability and folding like their capability to be post-translationally changed. Since histones are the most chromatin proteins, any alteration in abundance, post-transnational modification (PTMs) or structure will have acute effect on the chromatin global structure, gene expression effectiveness, genome replication and stability, several reports are also clarified that OS promote alteration modified in histones methylation and acetylation as it represents over the maintain the chromatin state enzymes. OS generate via H₂O₂ can induce histone alteration complexes to inhibit active tumor suppressor genes. In spite of OS impacting histones posttranslational modifications which involved in chromatin regulation, it does not effect in the similar manner according to the variant sensibility to the OS of the Histone Methyltransferases, HDMs and HATs.

In a same way to DNAM, one of the notable alteration is histone deacetylases (HDAC), that eliminate their activity by cigarette smoke^[65]. The patients with COPD demonstrate lower level of HDAC2 activity that raised acetylation in histones H3 and H4 of the NF-κB promoter therefore pro-inflammatory genes dys-regulation^[66].

Oxidative stress Effect on Non-Coding RNAs: The regulation of transcription depended on non-coding RNAs is also change in diffident manner by OS, as it has demonstrated in modifications of histones and DNA. To yet, there are several microRNAs whose their generation changes due to cellular OS changing^[67,68]. like, in epithelial cells miR-200c is up-regulated as consequences of increasing ROS that lead to elevate senescent cells and apoptotic via the its target gene ZEB1action^[69]. some miRNAs is stimulate by transcription factors that are sensitive to elevated ROS level like miR-27a/b miR-506, miR-200 and miR-206 via its target genes^[70,71-73]. furthermore, the miRNAs processing from primary structure is regulated by DGRC8-Drosha complexes which eliminated the processing capacity of DGRC8 by OS, that observed on Fe(III) for its action and therefore corresponding mature miRNA is down regulated^[74].

some miRNAs implicated in the detoxification pathway of NRF2-ARE, according to its natural inhibitor KEAP1 or the targeting of NRF2, or indirect action of genes which regulate the previous signaling pathway, like, NRF2 expression is inhibited by miR-101 in breast cancer cells to activate its sensitivity to suppress proliferation and OS^[75]. in esophageal squamous cell carcinoma miR-432-3p directly linked to the coding region of KEAP1 to down regulating and transcription of downstream genes up-regulating of the NRF2-ARE

pathway^[76]. In neuroblastoma cells miR-7 relieves the OS via targeting KEAP1^[77]. in different cancer cell miR-200a linked to the 3' UTR sequence of KEAP1 causing degradation of its mRNA^[78-80].

On the other hand, Several miRNAs Expression have been clarified to be changes under OS, Magenta in vitro experiment have estimated ROS impact in the miRNAs levels in the endothelial cells. Study found that human umbilical vein endothelial cells Exposure to H₂O₂ causes significant miR-200c and miR-141 up-regulation. Yu^[81] have clarified the miR-200c role in the ROS-associated apoptosis regulation.

miR-21 is one of the miRNA types with role in the response to OS which miRNA has been found to be stimulated via ultraviolet b in cell line via ROS-mediated MAPK pathways^[82]. Conversely, human umbilical vein endothelial cells miR-21 has proved to ROS homeostasis interrupted and antioxidant responses disturbance in cellular glucose variability^[83]. miR-145 is up-regulated through OS is. Moreover, lncRNAs have been found to be associated with OS, like level of cancer-associated lncRNAs were increased in the some disease like myocardial infarction.

CONCLUSIONS

According to the information mentioned above, oxidative stress effect in chromatin at different levels included DNA, histones, structural DNA-binding proteins and histone modifiers. As well-known that OS is implicated in disease pathogenicity. However, belong to the mechanisms of epigenetic affected by OS, the direct association of OS in disease has not demonstrated, the chromatin alterations mediated by ROS have significant impacts in initiation and progression of some diseases. From these literature review can be concluded the significant ROS role in DNAM included global hypo methylation and specific gene hypermethylation and methylation enzymes, regarding to histone, its affected by ROS or NOS at methylation acetylation and phosphorylation. Oxidative stress also effected and affected by non-coding RNA included microRNA and lncRNAs in addition to other types of RNA, other epigenetic mechanisms also undergo the OS changes. Further investigation need to be implemented for more information's particularly in common diseases.

REFERENCES

1. Bishop, K. and L. Ferguson, 2015. The interaction between epigenetics, nutrition and the development of cancer. *Nutrients*, 7: 922-947.
2. Stirzaker, C., P.C. Taberlay, A.L. Statham and S.J.

- Clark, 2014. Mining cancer methylomes: Prospects and challenges. *Trends Genet.*, 30: 75-84.
3. Gomez, M., J. Wu, V. Schreiber, J. Dunlap, F. Dantzer, Y. Wang and Y. Liu, 2006. Parp1 is a trf2-associated poly(ADP-ribose)polymerase and protects eroded telomeres. *Mol. Biol. Cell*, 17: 1686-1696.
 4. Zampieri, M., F. Ciccarone, R. Calabrese, C. Franceschi, A. Bürkle and P. Caiafa, 2015. Reconfiguration of DNA methylation in aging. *Mech. Ageing Dev.*, 151: 60-70.
 5. Deplus, R., 2002. Dnmt3l is a transcriptional repressor that recruits histone deacetylase. *Nucleic Acids Res.*, 30: 3831-3838.
 6. Brown, S.D.M., 1991. Xist and the mapping of the x chromosome inactivation centre. *BioEssays*, 13: 607-612.
 7. Ng, K., D. Pullirsch, M. Leeb and A. Wutz, 2007. Xist and the order of silencing. *EMBO reports*, 8: 34-39.
 8. Boumil R.M, Y. Ogawa, B.K. Sun, K.D Huynh, J.T. Lee. 2006. Differential methylation of Xite and CTCF sites in Tsix mirrors the pattern of X-inactivation choice in mice. *Mol. Cell. Biol.* 26: 2109-2117.
 9. Reik, W. and J. Walter, 2001. Genomic imprinting: Parental influence on the genome. *Nat. Rev. Genet.*, 2: 21-32.
 10. Portela, A. and M. Esteller, 2010. Epigenetic modifications and human disease. *Nat. Biotechnol.*, 28: 1057-1068.
 11. Bannister, A.J. and T. Kouzarides, 2011. Regulation of chromatin by histone modifications. *Cell Res.*, 21: 381-395.
 12. Yang, W., Y. Xia, D. Hawke, X. Li and J. Liang *et al.*, 2012. Pkm2 phosphorylates histone h3 and promotes gene transcription and tumorigenesis. *Cell*, 150: 685-696.
 13. Lau, P.N.I. and P. Cheung, 2011. Histone code pathway involving h3 s28 phosphorylation and k27 acetylation activates transcription and antagonizes polycomb silencing. *Proc. Nat. Acad. Sci.*, 108: 2801-2806.
 14. Dawson, M.A., A.J. Bannister, B. Göttgens, S.D. Foster, T. Bartke, A.R. Green and T. Kouzarides, 2009. Jak2 phosphorylates histone h3y41 and excludes hp1a from chromatin. *Nature*, 461: 819-822.
 15. Cohen-Armon, M., L. Visochek, D. Rozensal, A. Kalal and I. Geistrikh *et al.*, 2007. Dna-independent parp-1 activation by phosphorylated erk2 increases elk1 activity: A link to histone acetylation. *Mol. Cell*, 25: 297-308.
 16. Moss, E. and M.E. Halkos, 2017. Cost effectiveness of robotic mitral valve surgery. *Ann. Cardiothorac. Surg.*, 6: 33-37.
 17. Gill, G., 2005. Something about sumo inhibits transcription. *Curr. Opin. Genet. And Dev.*, 15: 536-541.
 18. Kshetrimayum, C., A. Sharma, V.V. Mishra and S. Kumar, 2019. Polycystic ovarian syndrome: Environmental/occupational, lifestyle factors: An overview. *J. Turk. German Gynecol. Assoc.*, 20: 255-263.
 19. Meas, R. and P. Mao, 2015. Histone ubiquitylation and its roles in transcription and DNA damage response. *DNA Repair*, 36: 36-42.
 20. Oya, E., R. Nakagawa, Y. Yoshimura, M. Tanaka and G. Nishibuchi *et al.*, 2019. H3k14 ubiquitylation promotes h3k9 methylation for heterochromatin assembly. *EMBO reports*, Vol. 20 .10.15252/embr.201948111.
 21. Kim, J., M. Guermah, R.K. McGinty, J.S. Lee and Z. Tang *et al.*, 2009. Rad6-mediated transcription-coupled h2b ubiquitylation directly stimulates h3k4 methylation in human cells. *Cell*, 137: 459-471.
 22. Mattick, J.S. and I.V. Makunin, 2006. Non-coding rna. *Hum. Mol. Genet.*, 15: 17-29.
 23. Gibb, E.A., C.J. Brown and W.L. Lam, 2011. The functional role of long non-coding RNA in human carcinomas. *Mol. Cancer*, Vol. 10 .10.1186/1476-4598-10-38.
 24. Djebali, S., C.A. Davis, A. Merkel, A. Dobin and T. Lassmann *et al.*, 2012. Landscape of transcription in human cells. *Nature*, 489: 101-108.
 25. Advani, K., M. Batra, S. Tajpuriya, R. Gupta and A. Saraswat *et al.*, 2019. Efficacy of combination therapy of inositols, antioxidants and vitamins in obese and non-obese women with polycystic ovary syndrome: An observational study. *J. Obstet. Gynaecol.*, 40: 96-101.
 26. Waddington, C.H., 1939. Preliminary notes on the development of the wings in normal and mutant strains of drosophila. *Proc. Nat. Acad. Sci.*, 25: 299-307.
 27. Holliday, R., 1987. The inheritance of epigenetic defects. *Science*, 238: 163-170.
 28. Deans, C. and K.A. Maggert, 2015. What do you mean, "epigenetic" *Genetics*, 199: 887-896.
 29. Egger, G., G. Liang, A. Aparicio and P.A. Jones, 2004. Epigenetics in human disease and prospects for epigenetic therapy. *Nature*, 429: 457-463.
 30. Sharma, S., T.K. Kelly and P.A. Jones, 2009. Epigenetics in cancer. *Carcinogenesis*, 31: 27-36.
 31. Wu, H. and Y. Zhang, 2014. Reversing DNA methylation: Mechanisms, genomics and biological functions. *Cell*, 156: 45-68.
 32. Oda, M., D. Oxley, W. Dean and W. Reik, 2013.

- Regulation of lineage specific DNA hypomethylation in mouse trophoblast. PLoS ONE, Vol. 8 .10.1371/journal.pone.0068846.
33. Jurkowska, R.Z., T.P. Jurkowski and A. Jeltsch, 2010. Structure and function of mammalian DNA methyltransferases. *ChemBioChem*, 12: 206-222.
 34. Bostick, M., J.K. Kim, P.O. Estève, A. Clark, S. Pradhan and S.E. Jacobsen, 2007. Uhrf1 plays a role in maintaining DNA methylation in mammalian cells. *Science*, 317: 1760-1764.
 35. Kohli, R.M. and Y. Zhang, 2013. Tet enzymes, tdg and the dynamics of DNA demethylation. *Nature*, 502: 472-479.
 36. Nabel, C.S., H. Jia, Y. Ye, L. Shen and H.L. Goldschmidt *et al.*, 2012. Aid/apobec deaminases disfavor modified cytosines implicated in DNA demethylation. *Nat. Chem. Biol.*, 8: 751-758.
 37. Dasuri, K., L. Zhang and J.N. Keller, 2013. Oxidative stress, neurodegeneration and the balance of protein degradation and protein synthesis. *Free Radical Biol. Med.*, 62: 170-185.
 37. Newsholme, P., E. Rebelato, F. Abdulkader, M. Krause, A. Carpinelli and R. Curi, 2012. Reactive oxygen and nitrogen species generation, antioxidant defenses, and β -cell function: A critical role for amino acids. *J. Endocrinol.*, 214: 11-20.
 38. Rigoulet, M., E.D. Yoboue and A. Devin, 2011. Mitochondrial ros generation and its regulation: Mechanisms involved in h2O2 signaling. *Antioxidants & Redox Signaling*, 14: 459-468.
 39. Murphy, M.P., 2008. How mitochondria produce reactive oxygen species. *Biochem. J.*, 417: 1-13.
 40. Nagaria, T., A. Mohapatra and J. Jaiswal, 2019. Effect of myoinositol and metformin in combination on clinical and hormonal profile in patients of polycystic ovarian syndrome. *Int. J. Reprod. Contracep. Obstet. Gynecol.*, 8: 702-709.
 41. Cencioni, C., F. Spallotta, F. Martelli, S. Valente, A. Mai, A. Zeiher and C. Gaetano, 2013. Oxidative stress and epigenetic regulation in ageing and age-related diseases. *Int. J. Mol. Sci.*, 14: 17643-17663.
 42. Lewandowska, J. and A. Bartoszek, 2011. Dna methylation in cancer development, diagnosis and therapy--multiple opportunities for genotoxic agents to act as methylome disruptors or remediators. *Mutagenesis*, 26: 475-487.
 43. Chia, N., L. Wang, X. Lu, M.C. Senut, C.A. Brenner and D.M. Ruden, 2011. Hypothesis: Environmental regulation of 5-hydroxymethylcytosine by oxidative stress. *Epigenetics*, 6: 853-856.
 44. Simpson, N.E., V.P. Tryndyak, M. Pogribna, F.A. Beland and I.P. Pogribny, 2012. Modifying metabolically sensitive histone marks by inhibiting glutamine metabolism affects gene expression and alters cancer cell phenotype. *Epigenetics*, 7: 1413-1420.
 45. Nishida, N. and M. Kudo, 2013. Oxidative stress and epigenetic instability in human hepatocarcinogenesis. *Digestive Dis.*, 31: 447-453.
 46. Pundir, C., R. Deswal, V. Narwal and A. Dang, 2020. The prevalence of polycystic ovary syndrome: A brief systematic review. *J. Hum. Reprod. Sci.*, 13: 261-271.
 47. Kreuz, S. and W. Fischle, 2016. Oxidative stress signaling to chromatin in health and disease. *Epigenomics*, 8: 843-862.
 48. Nishida, N., T. Arizumi, M. Takita, S. Kitai and N. Yada *et al.*, 2013. Reactive oxygen species induce epigenetic instability through the formation of 8-hydroxydeoxyguanosine in human hepatocarcinogenesis. *Digestive Dis.*, 31: 459-466.
 49. Udomsinprasert, W., N. Kitkumthorn, A. Mutirangura, V. Chongsrisawat, Y. Poovorawan and S. Honsawek, 2016. Global methylation, oxidative stress and relative telomere length in biliary atresia patients. *Sci. Rep.*, Vol. 6 .10.1038/srep26969.
 50. Ziech, D., R. Franco, A. Pappa and M.I. Panayiotidis, 2011. Reactive oxygen species (ros)—induced genetic and epigenetic alterations in human carcinogenesis. *Mutat. Res./Fundam. Mol. Mech. Mutagen.*, 711: 167-173.
 51. Deaton, A.M. and A. Bird, 2011. CpG islands and the regulation of transcription. *Genes and Dev.*, 25: 1010-1022.
 52. Dickson, K.M., C.B. Gustafson, J.I. Young, S. Züchner and G. Wang, 2013. Ascorbate-induced generation of 5-hydroxymethylcytosine is unaffected by varying levels of iron and 2-oxoglutarate. *Biochem. Biophys. Res. Commun.*, 439: 522-527.
 53. Ponnaluri, V.K.C., J.P. Maciejewski and M. Mukherji, 2013. A mechanistic overview of tet-mediated 5-methylcytosine oxidation. *Biochem. Biophys. Res. Commun.*, 436: 115-120.
 54. Mastroeni, D., A. Grover, E. Delvaux, C. Whiteside, P.D. Coleman and J. Rogers, 2010. Epigenetic changes in alzheimer's disease: Decrements in DNA methylation. *Neurobiology Aging*, 31: 2025-2037.
 55. Volkmar, M., S. Dedeurwaerder, D.A. Cunha, M.N. Ndlovu and M. Defrance *et al.*, 2012. Dna methylation profiling identifies epigenetic dysregulation in pancreatic islets from type 2 diabetic patients. *The EMBO J.*, 31: 1405-1426.
 56. Cheishvili, D., B. Stefanska, C. Yi, C.C. Li and P. Yu *et al.*, 2015. A common promoter hypomethylation signature in invasive breast, liver and prostate

- cancer cell lines reveals novel targets involved in cancer invasiveness. *Oncotarget*, 6: 33253-33268.
57. Leodolter, A., S. Alonso, B. González, M.P. Ebert and M. Vieth *et al.*, 2015. Somatic DNA hypomethylation in *H. pylori* -associated high-risk gastritis and gastric cancer: Enhanced somatic hypomethylation associates with advanced stage cancer. *Clin. Transl. Gastroenterol.*, Vol. 6 .10.1038/ctg.2015.14.
 58. GRAVINA, G.L., G. RANIERI, P. MUZI, F. MARAMPON and A. MANCINI *et al.*, 2012. Increased levels of DNA methyltransferases are associated with the tumorigenic capacity of prostate cancer cells. *Oncol. Rep.*, 29: 1189-1195.
 59. Niu, Y., T.L. DesMarais, Z. Tong, Y. Yao and M. Costa, 2015. Oxidative stress alters global histone modification and DNA methylation. *Free Radical Biol. Med.*, 82: 22-28.
 60. Delatte, B., J. Jeschke, M. Defrance, M. Bachman and C. Creppe *et al.*, 2015. Genome-wide hydroxymethylcytosine pattern changes in response to oxidative stress. *Sci. Rep.*, Vol. 5 .10.1038/srep12714.
 61. Xin, Y.J., B. Yuan, B. Yu, Y.Q. Wang, J.J. Wu, W.H. Zhou and Z. Qiu, 2015. Tet1-mediated DNA demethylation regulates neuronal cell death induced by oxidative stress. *Sci. Rep.*, Vol. 5 .10.1038/srep07645.
 62. O'Hagan, H.M., W. Wang, S. Sen, C.D. Shields and S.S. Lee *et al.*, 2011. Oxidative damage targets complexes containing DNA methyltransferases, sirt1, and polycomb members to promoter cpG islands. *Cancer Cell*, 20: 606-619.
 63. Ding, N., E.M. Bonham, B.E. Hannon, T.R. Amick, S.B. Baylin and H.M. O'Hagan, 2016. Mismatch repair proteins recruit DNA methyltransferase 1 to sites of oxidative DNA damage. *J. Mol. Cell Biol.*, 8: 244-254.
 64. Yao, H. and I. Rahman, 2012. Role of histone deacetylase 2 in epigenetics and cellular senescence: Implications in lung inflammaging and copd. *Am. J. Physiol.-Lung Cell. Mol. Physiol.*, 303:
 65. Arit, A., 2014. Oxidative stress and the epigenome in human disease. *J. Genet. Genome Res.*, Vol. 1 .10.23937/2378-3648/1410010.
 66. Fuschi, P., B. Maimone, C. Gaetano and F. Martelli, 2019. Noncoding rnas in the vascular system response to oxidative stress. *Antioxidants and Redox Signaling*, 30: 992-1010.
 67. Lan, J., Z. Huang, J. Han, J. Shao and C. Huang, 2018. Redox regulation of micrnas in cancer. *Cancer Lett.*, 418: 250-259.
 68. Magenta, A., C. Cencioni, P. Fasanaro, G. Zaccagnini and S. Greco *et al.*, 2011. Mir-200c is upregulated by oxidative stress and induces endothelial cell apoptosis and senescence via zeb1 inhibition. *Cell Death and Differentiation*, 18: 1628-1639.
 69. Yang, H., T.W.H. Li, Y. Zhou, H. Peng and T. Liu *et al.*, 2015. Activation of a novel c-myc-mir27-prohibitin 1 circuitry in cholestatic liver injury inhibits glutathione synthesis in mice. *Antioxidants & Redox Signaling*, 22: 259-274.
 70. Xiao, Y., W. Yan, L. Lu, Y. Wang, W. Lu, Y. Cao and W. Cai, 2015. Retracted article: P38/p53/mir-200a-3p feedback loop promotes oxidative stress-mediated liver cell death. *Cell Cycle*, 14: 1548-1558.
 71. Yin, M., X. Ren, X. Zhang, Y. Luo and G. Wang *et al.*, 2014. Selective killing of lung cancer cells by miRNA-506 molecule through inhibiting nf-b p65 to evoke reactive oxygen species generation and p53 activation. *Oncogene*, 34: 691-703.
 72. Faller, M., M. Matsunaga, S. Yin, J.A. Loo and F. Guo, 2006. Heme is involved in microrna processing. *Nat. Struct. And Mol. Biol.*, 14: 23-29.
 73. Barr, I., A.T. Smith, Y. Chen, R. Senturia, J.N. Burstyn and F. Guo, 2012. Ferric, not ferrous, heme activates rna-binding protein dgcr8 for primary microrna processing. *Proc. Nat. Acad. Sci.*, 109: 1919-1924.
 74. Muñoz, J.S.G., D.J. Rodríguez and J.J.H. Morante, 2014. Diurnal rhythms of plasma GLP-1 levels in normal and overweight/obese subjects: Lack of effect of weight loss. *J. Physiol. Biochem.*, 71: 17-28.
 75. Akdemir, B., Y. Nakajima, J. Inazawa and J. Inoue, 2017. Mir-432 induces nrf2 stabilization by directly targeting keap1. *Mol. Cancer Res.*, 15: 1570-1578.
 76. Kabaria S., D.C.Choi., A.D.Chaudhuri ., M.R.Jain ., H.Li and E.Junn . 2015. MicroRNA-7 activates Nrf2 pathway by targeting Keap1 expression *Free Radic. Biol. Med* 89: 548-556.
 77. Eades, G., M.Yang ., Y.Yao ., Y.Zhang .and Zhou Q 2011. miR-200a regulates Nrf2 activation by targeting Keap1 mRNA in breast cancer cells *J. Biol. Chem.* 286: 40725-40733.
 78. Petrelli, A., A. Perra, D. Cora, P. Sulas and S. Menegon *et al.*, 2013. Microrna/gene profiling unveils early molecular changes and nuclear factor erythroid related factor 2 (nrf2) activation in a rat model recapitulating human hepatocellular carcinoma (hcc). *Hepatology*, 59: 228-241.
 79. Liu, M., C.Hu., Q.Xu ., L.Chen ., K.Ma ., N.Xu and H.Zhu 2015. Methylseleninic acid activates Keap1/Nrf2 pathway via up-regulating miR-200a in human oesophageal squamous cell carcinoma cells *Biosci. Rep.* Vol. 35 .10.1042/BSR20150092.
 80. Yu, D.S., G. Lv, X.F. Mei, Y. Cao, Y.F. Wang, Y.S. Wang and Y.L. Bi, 2014. Mir-200c regulates ros-induced

- apoptosis in murine bv-2 cells by targeting fap-1. Spinal Cord, 53: 182-189.
81. Hou, L., L. Bowman, T.G. Meighan, P. Pratheeshkumar, X. Shi and M. Ding, 2013. Induction of mir-21-pdcd4 signaling by uvb in jb6 cells involves ros-mediated mapk pathways. Exp. Toxicol. Pathol., 65: 1145-1148.
82. Sala, L.L., S. Mrakic-Sposta, S. Micheloni, F. Prattichizzo and A. Ceriello, 2018. Glucose-sensing microrna-21 disrupts ros homeostasis and impairs antioxidant responses in cellular glucose variability. Cardiovasc. Diabetology, Vol. 17 .10.1186/s12933-018-0748-2.